INTRODUCTION

Hyperlipidemia and hypercholesterolemia are major biochemical risk factors for atherogenesis, which ultimately contribute to atherothrombosis with severe consequences. Currently, there is a small number of research articles dedicated to the structural changes in the aorta caused by hyperlipidemia and atherosclerosis as well as HSV-induced atherosclerosis [1]. Even less information is available on morphological studies of the aorta affected by a viral infection associated with atherogenesis. HSV and CMV were detected in 40% of the examined autopsy specimens of the aorta and coronary vessels [2]. In experiments, the effect of HSV-infection on atherosclerosis has been studied fragmentarily, but nevertheless they demonstrate the formation of atheroma in mice with murine gammaherpesvirus-68 (MHV-68) and its progression within 24 weeks compared to the control group [3]. Murine CMV (MCMV) increases the incidence rate of atherothrombosis in apoE-knockout mice by 84%, whereas Chlamydia pneumoniae (Cpn) are not associated with an increased rate of the disease [4]. Morphological signs of atherosclerosis are described in the aortic sinus in apoE-knockout mice [5]. The fact that HSV-1-induced atherothrombosis does not reach the values of atheroma is linked to the development of systemic infection, even though HSV-infection promotes thrombosis and increases the risk of atherothrombosis [6].

It is currently widely recognized that any viral infection that damages endothelial cells of blood vessels can be reliably associated with the risk of vascular disease [7]. Biochemical analysis revealed that content of free and esterified cholesterol was higher in the aortic tissue specimens of CMV-infected animals with normocholesterolemia than in the models of non-infected animals. Thus, according to Fabricant C., herpesvirus infection contributed to the accumulation of lipids in the aorta [8], and previous immunization prevented atherosclerotic and fibroproliferative changes in the aortic wall [9] due to decreased activity of cytoplasmic cholesterol esterase in animals [10]. The findings were consistent with accumulation of saturated cholesterol and triacylglycerol due to impaired hydrolysis observed in the HSV-1-infected smooth muscle cells in humans [11]. The scholars describe the smooth muscle cells involved in the active synthesis of extracellular matrix of the connective tissue, such as glycosaminoglycans, collagen and elastin [12]. It is believed that the described cells originate from the media, but penetrate into the intima through the damaged inner elastic membrane of the aorta, and these altering changes are an early manifestation of atherogenesis in the intima of hypercholesterolemic animals [13].

Our hypothesis is that previous viral infection induces structural and functional changes in the tunica intima of
blood vessels that may promote the accumulation of lipid inclusions in the aortic intima in animals consuming a high-fat and high-cholesterol diet.

THE AIM
To study the effect of a high-fat diet (HFD) on the structural changes in the aortic intima in intact and HSV-1-infected mice using Scanning electron microscopy (SEM) and Transmission electron microscopy (TEM).

MATERIALS AND METHODS
Balb/c mice were used in the experiments considered here. The study included 5 groups of the murine aortic tissue samples:

Control group (n = 3) – intact animals;

Group 1 – HSV-1-infected animals (n = 4);

Group 2 – animals consuming high-fat diet (HFD) (n = 6);

Group 3 – HSV-1-infected animals that were subsequently consuming a high-fat diet (HSV / HFD) (n = 6);

Group 4 – animals consuming a high-fat diet that were subsequently infected with HSV-1 (HFD / HSV) (n = 6). The animals in the 1st group were infected with HSV-1 in L.V. Gromashhevsky Institute of Epidemiology and Infectious Diseases, NAMS of Ukraine (Head of the Laboratory of Experimental Chemotherapy of Viral Infections, MD, Prof. S.L. Rybalko).

In a 12-week experiment, the animals in the 2nd group received the food with cholesterol, animal and vegetable fats. 100 g of food contained 30-45 g of heat-treated animal fats (pork fat / lard, butter); 15-20 g of heat-treated vegetable fats (margarine); 1.5-2 g of cholesterol; 1 g of calcium carbonate (CaCO₃); 10 mg (in terms of daily intake for animals – 10 mg / kg) of Mercazolyl (1-methyl 2-mercaptoimidazole with potato starch, Calcium stearate, refined sugar, talcum), 32-52 g of standard granulated animal feed.

The animals in the 3rd group were infected with HSV-1 and then received a high-fat diet as above.

The animals in the 4th group were not analyzed because the group had an absolute mortality in the first 3-5 days after infection.

RESUL TS

High-magnification SEM images (×8000 – ×13000) showed a longitudinal waviness of the aortic lumen in intact animals, which is physiological. In the 1st (HSV), 2nd (HFD) and 3rd (HSV / HFD) groups, the surface had the same physiological waviness that correlates with the presence of the inner elastic membrane in the aorta and indicates its preservation.

Low-magnification SEM images (×1100 – ×1500) showed a longitudinal waviness of the aortic lumen in intact animals, which is physiological. In the 1st (HSV), 2nd (HFD) and 3rd (HSV / HFD) groups, the surface had the same physiological waviness that correlates with the presence of the inner elastic membrane in the aorta and indicates its preservation.
protrusions in the 2nd (HFD) group (Fig. 1). This is due to edema and elevation of endothelial cells, their nuclear zone, swelling of cytoplasm above the basal membrane. The accumulation of extracellular matrix derivates and necrotic cells in the subendothelial space is not excluded.

In the 1st (HSV) and 3rd (HSV / HFD) groups, SEM showed adhesion of leukocytes on the surface of the aortic intima in all areas. Additionally, TEM registered leukocytes in the subendothelial zone, which can be explained by their penetration through the endothelial layer. There were no gaps in the endothelium and platelet adhesion detected in the 1st (HSV) and 2nd (HFD) groups, whereas in the 3rd (HSV / HFD) group, we recorded necrotized endothelial cells, signs of coagulation and loss of intimal integrity.

The surface of the intima had additional signs of destructive changes in the form of fibrous structures that differed from the intact aorta. Those were damaged endothelial cells and most likely coagulated proteins. Such changes were more prominent in the 3rd (HSV / HFD) than in the 1st (HSV) group (Fig. 1).

On the surface of the intima in the 2nd (HFD) group, erythrocyte adhesion and coagulation products were observed, but leukocytes were found less frequently. All groups had focal zones of disorganization and loss of endothelial lining (disorganization), which distinguished experimental samples from intact aortic specimens. According to SEM, erythrocyte adhesion and endothelial cell edema, seen as focal protrusions on the surface, were denser in the 2nd (HFD) than in the 1st (HSV) and 3rd (HSV / HFD) groups (unfortunately, quantitative analysis is impossible). Damage and loss of endothelial cells in the intima were more pronounced in the 1st (HSV) and 3rd (HSV / HFD) groups than in the 2nd (HFD) group due to the presence of a viral infection, which can explain more severe necrotic and lytic changes in endothelial cells (with the formation of focal regions, where endothelium is irregular or absent).

TEM was conducted to evaluate the following structural elements of the aortic wall: 1) endothelial layer; 2) subendothelial zone; 3) muscle membrane. In the control group, the aortic intima was unchanged: the nuclei of endothelial cells had a different distribution of chromatin, cytoplasmic...
Table 1. Results of quantitative measurements in comparison groups (Me [Q1-Q3])

<table>
<thead>
<tr>
<th>Group</th>
<th>Infectious titer, Ig CPE 50</th>
<th>Diameter of lipid inclusions, micron</th>
<th>Diameter of atypical vesicles in endothelial cells, micron</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Group 1 (HFD)</td>
<td>-</td>
<td>0,48[0,30-0,68]</td>
<td>0,38[0,23-0,47]</td>
</tr>
<tr>
<td>Group 2 (HSV)</td>
<td>1[1-2]</td>
<td>-</td>
<td>0,29[0,19-0,38]</td>
</tr>
<tr>
<td>Group 3 (HSV/HFD)</td>
<td>2[1-3]</td>
<td>0,66[0,46-0,98]*</td>
<td>0,68[0,43-0,67]*#</td>
</tr>
<tr>
<td>Group 4 (HFD/HSV)</td>
<td>3,5[2,75-4]#</td>
<td>NA</td>
<td>NA</td>
</tr>
</tbody>
</table>

Note: * - significant difference with the 2nd (HFD) group; # - significant difference with the 1st (HSV) group; NA - not analyzed

pinocytotic vesicles; in smooth muscle cells, intact organelles (ribosomes, vesicles, mitochondria, endoplasmic reticulum) were found (Fig. 2).

In the 2nd (HFD) group, SEM showed organelle reduction, edema and destruction of mitochondria, deformation of the nuclei in endothelial cells. Endothelium atypically protrudes into the lumen of vessel and separates the derivatives of cell destruction into the aortic lumen, which explains the appearance of the altered intimal contour. In some cells pinocytotic vesicles were found (the evidence of partial preservation of transendothelial transport). In the endothelium cytoplasm, there were detected atypical vesicles, most likely formed as a result of swelling of membrane organelles, in particular mitochondria. Their diameter was evaluated morphometrically (Table 1).

The inner elastic membrane of the aorta was structurally preserved. In the subendothelial zone of the aorta, lipid inclusions were detected. The diameter of lipid inclusions in macrophages was in the range of 150-230 nm, and in the extracellular matrix it even reached 4 microns (Table 1).

Lipid deposition contributed to the transformation of cholesterol-engorged macrophages into so-called «foam cells», destruction of individual cells and the appearance of cell free zones. Some cells were in a state of total destruction.

The morphological organization and the integrity of the murine aortic muscle membrane were preserved in the 2nd (HFD) group, even though some changes in smooth muscle cells were found: 1) edema and destruction of cristae of mitochondria, 2) appearance of atypical vesicles in the cytoplasm, and 3) reduction of organelles and myofibrils. In some cells, accumulation of secondary lysosomes was observed. These changes indicate a secondary metabolic injury.

In endothelial cells of the 1st (HSV) group, the following changes were recorded: 1) cytoplasmic edema; 2) lysis of organelles; 3) atypical vesicles, which can be considered as a consequence of lytic changes and fusion of membrane organelles. In this case, the integrity of the endothelial layer was preserved. Under the endothelium, the basal membrane, internal elastic membrane, and subendothelial layer of the inner aortic membrane were observed. In subendothelial zone, the following changes were revealed: 1) macrophages; 2) diapedesis of some blood corpuscles, 3) decrease in the density of collagen fibers. The revealed changes indicate a decrease in: 1) the barrier function of endothelial cells, 2) the infiltration of leukocytes; 3) the reorganization of paravasal intercellular space, which is a manifestation of the inflammatory reaction and is a qualitative morphological distinction from the 2nd (HFD) group. Under the elastic membrane, smooth muscle cells of the aortic media were found without any significant structural changes.

The 3rd (HSV / HFD) group had progressive structural changes, as described in the 1st (HFD) group. Changes in the endothelium in the form of: 1) edema and destruction of organelles, 2) the appearance of atypical vesicles of greater diameter; 3) necrosis of cells with desquamation and endothelial damage. During SEM these changes were seen as: 1) sites of destruction, coagulation; 2) loss of the endothelial cells (Table 1). The increase in the density and diameter of atypical vesicles by 57.8% (P = 0.05) indicates the progression of dystrophic changes in the aortic endothelial cells in the 3rd (HSV / HFD) group compared to the 2nd (HFD) group. Accumulation of virions (diameter 120-150 nm) in injured cell was detected in the one sample.

In the subendothelial layer, the following changes were also revealed: 1) accumulation of lipid inclusions, 2) destruative changes, 3) reorganization of the extracellular matrix and infiltration of macrophages. The diameter of lipid inclusions was 37.5% higher (P <0.05) compared to the 2nd (HFD) group. The density of macrophages with «foamy» cytoplasm also increased, and the reorganization of collagen fibers can be explained by phagocytic activity of macrophages and their participation in remodeling of extracellular matrix. This indicates a higher degree of structural damage to the aorta in the 3rd (HSV / HFD) group compared to the 2nd (HFD) group, that is HSV1 potentiates their development.

The morphological integrity of the muscle membrane was preserved. But, whereas in the 2nd (HFD) group only atypical vesicles were detected in smooth muscle cells, in the 3rd (HSV / HFD) group, perinuclear edema of the nucleus was additionally revealed. This indicates that the presence of HSV infection potentiates the development of ultrastructural changes in the aortic media. Changes in smooth muscle cells are nonspecific and most likely related to metabolic disorders caused by a 12-week consumption of a high-fat diet in the 2nd (HFD) group.

TEM demonstrated that injured aortic intima had insufficient number of ultrastructural markers to identify and verify the damage factor. Lipid inclusions occurred
in the 2\textsuperscript{nd} (HFD) and 3\textsuperscript{rd} (HSV / HFD) groups, and signs of viral damage to endothelial cells (lamellar cells, single virions) were detected only in one sample from the 3\textsuperscript{rd} (HSV / HFD) group.

In vitro studies in all cardiac homogenates confirmed the presence of HSV-1, and the level of reproduction was estimated according to CPE (Table 1). Although there was no significant difference between the 1\textsuperscript{st} (HSV) and the 3\textsuperscript{rd} (HSV / HFD) groups, fatal cases of another model of the 4\textsuperscript{th} group (HFD / HSV) caused significantly higher CPE in the hearts. It is clear that the term of infection period is different, but the absolute lethality of mice that received a high-fat diet for 12 weeks and then were infected with HSV-1, gives reason to think about a certain direct and inverse relationship in the study.

**DISCUSSION**

Changes in endothelial cells and extracellular matrix were more pronounced in the 3\textsuperscript{rd} (HSV / HFD) group compared to the 2\textsuperscript{nd} (HFD) group. The most striking changes in the aortic intima were observed as the accumulation of lipid inclusions and macrophages in the subendothelial space. The revealed changes in the subendothelial space of the aorta can be explained by: 1) migration of monocytes / macrophages from blood or regional phagocytes for the elimination of lipid inclusions in the 2\textsuperscript{nd} (HFD) group; 2) adhesion and migration of leukocytes to the vascular wall for aggressive response to HSV-1 in the 1\textsuperscript{st} (HSV) group; 3) lipid deposition of cellular debris in the extracellular matrix and macrophages (lysosomal deposits) promoted by a structurally damaged endothelial layer under the influence of HSV-1 in the 3\textsuperscript{rd} (HSV / HFD) group. The lipid inclusions and necrotic cell debris were phagocyted by macrophages in subendothelial space of the 2\textsuperscript{nd} (HFD) and 3\textsuperscript{rd} (HSV / HFD) groups. However, these processes were more intense in the 3\textsuperscript{rd} (HSV / HFD) group, the density of the detected phagocytes, the number and diameter of lysosomal inclusions were also greater in this group. In the 1\textsuperscript{st} (HSV) group, macrophages were also detected, but their migration, in our opinion, can be associated with the elimination of cellular detritus. The latter is due to lytic changes in the cells that are induced by HSV-1, and the appearance of individual lymphocytes within the aortic wall in response to the infection.

In the third group (HSV / HFD), damage to the sub-intimal smooth muscle cells (edema and destruction of mitochondria, decrease in electron density and fibrous structures) was detected. We believe that in the 3\textsuperscript{rd} (HSV / HFD) group ultrastructural changes progress from the aortic intima and subintimal matrix to the muscular layer. The prominent signs of necrosis of smooth muscle cells in mice after a 12-week high-fat diet were not observed (the damage was manifested only by the destruction of mitochondria).

SEM, used to evaluate the surface of the aorta, also yielded some interesting results. Prior to the experiment, we made an assumption about structural abnormalities in the aortic wall in the 2\textsuperscript{nd} (HFD) group that should have been in the form of changes on inner surface of aorta, but after 12 weeks of a high-fat diet those alterations did not occur. The aortic intima had a longitudinal physiological waviness similar to that of the intact aorta of mice, as described previously [14]. But other important peculiarities were observed in infected animals. The 1\textsuperscript{st} (HSV) and 3\textsuperscript{rd} (HSV / HFD) groups demonstrated «abundant» adhesion of leukocytes to the aortic intima, whereas the 2\textsuperscript{nd} (HFD) group showed more intense erythrocyte adhesion and swelling of the endothelial cells. Between those cells the structural defects were seen (gaps, lysis of cells with remnants of the nuclear zone of endotheliocytes). It was found that the contours of the neighboring cells did not have complete contact. The findings obtained by means of SEM correlate with TEM results and complement them. Similar gaps are described by other authors and are apparently a nonspecific manifestation of the aortic intima damage [15]. The results obtained in our study correlate with the findings that were published earlier.

**CONCLUSIONS**

1. The formation of atheroma in the aortic wall during a 12-week high-fat diet, as shown in the literature, was not detected.
2. Comparative analysis of the aortic changes in mice of the 1\textsuperscript{st}, 2\textsuperscript{nd} and 3\textsuperscript{rd} groups (HSV, HFD and HSV / HFD) proves the initiation of structural changes in the aortic intima including phagocytosis of individual lipid inclusions in the subendothelial space in the 2\textsuperscript{nd} (HFD) group and their subsequent accumulation in cells, cell damage and destruction, which leads to an increase in the density of lipid inclusions and cellular detritus in the extracellular aortic matrix in the 3\textsuperscript{rd} (HSV / HFD) group.
3. HSV infection potentiates the accumulation of lipid inclusions in the aortic intima during a HFD, facilitates infection and contributes to the development of acute infection.

**REFERENCES**


Research work performed by Bogomolets National Medical University: “Study of the features and consequences of stroke in patients of different age groups based on the genetic and infectious factors and comorbid pathology”

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Conflict of interest:
The Authors declare no conflict of interest.

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