

ORIGINAL ARTICLE
PRACA ORYGINALNA

PECULIARITIES OF THE EFFECTS OF BILE ACIDS ON ATPASE ACTIVITY OF THE COLON MUCOSA IN PATIENTS WITH OVERWEIGHT AND IRRITABLE BOWEL SYNDROME

DOI: 10.36740/WLek202003133

Iryna M. Ferents¹, Solomiia V. Bychkova², Mykola A. Bychkov¹

¹DANYLO HALYTSKYLVIV NATIONAL MEDICAL UNIVERSITY, LVIV, UKRAINE

²IVAN FRANKO NATIONAL UNIVERSITY, LVIV, UKRAINE

ABSTRACT

The aim is to investigate the effect of bile acids on the ATPase activity of the colon mucosa in patients with overweight and irritable bowel syndrome (IBS).

Materials and methods: Completely examined 12 patients with IBS and overweight. We estimated the ATPase activity of colon mucous of the patients with IBS spectrophotometrically by determined the content of orthophosphate that was released after ATP hydrolysis. We studied the effect of 3-sulphate of taurolithocholate (TLC-S) on specific activities of Na⁺/K⁺-ATPase, Ca²⁺-ATPase of endoplasmic reticulum (EPR), Ca²⁺-ATPase of plasmatic membrane (PM) and basal Mg²⁺-ATPase of postmitochondrial subcellular fraction of colon mucous of the patients with IBS.

Results: We established the specific activities of Na⁺/K⁺-ATPase, Ca²⁺-ATPase of EPR, Ca²⁺-ATPase of PM and basal Mg²⁺-ATPase. There were (6.06 ± 1.61), (5.88 ± 1.19), (8.86 ± 1.56) (6.44 ± 2.02) μmol P_i/mg protein per hour, respectively. TLC-S (50 μM) did not cause any change of Na⁺/K⁺-ATPase, as well as Ca²⁺-ATPase activities, but statistically significant increased activity of Mg²⁺-ATPase of postmitochondrial subcellular fraction of colon mucous of the patients with IBS by 4 fold.

Conclusions: TLC-S increased basal Mg²⁺-ATPase in the postmitochondrial fraction of colon mucous of the patients with overweight and IBS, but had no effect on Na⁺/K⁺-ATPase and Ca²⁺-ATPase activities. It has been suggested that activation of basal Mg²⁺-ATPase under by TLC-S may indicate the role of the endo-lysosomal system of epitheliocytes of colon mucous in developing of pathology IBS.

KEY WORDS: irritable bowel syndrome, overweight, ATPase, bile acids

Wiad Lek. 2020;73(3):574-577

INTRODUCTION

Increased food intake and a reduction in energy expenditure are responsible for the increase in excess body weight and subsequent obesity. Today, according to the World Health Organization, over one billion people are overweight on the planet. In Ukraine, approximately one third of the population has excess body weight [1]. Obesity is the cause of various somatic diseases, in particular, the gastrointestinal tract, including gastroduodenitis with nausea and functional vomiting and irritable bowel syndrome (IBS), which is most often associated with restrictive eating behavior. According to various authors, the combination of obesity with dyskinesias of the colon with constipation, diverticular disease, colon polyposis was diagnosed, respectively, at 36.28; 28.0 and 10.0% of patients. Other researchers have found that in obese individuals an association with functional constipation occurred in 24.0% of cases, and obesity was observed in 60.0% of patients with constipation [2].

Obesity also develops against a backdrop of stress, serving as an indicator of psycho-emotional maladaptation and overcoming difficult life situations that are inhibited by excessive eating. In addition, excess body weight can act as a factor that prevents pleasure from life, and the

latter phenomenon can be a factor that affects eating disorders, which in turn can contribute to the appearance of constipation, abdominal pain, changes in the sensitivity of serotonin receptors of the intestinal wall [2].

Obesity and a high body mass index have been shown to be significant risk factors for the development of IBS, in addition to insufficient amount of fiber in the diet, stress, inflammation, genetic predisposition [3]. Today, IBS is one of the most common diseases of the gastrointestinal tract, and obesity is an urgent problem of endocrinology [4]. IBS according to Rome Criteria IV is defined as a chronic functional bowel disorder characterized by recurrent abdominal pain, which occurs and continues at least once a week for the last three months, associated with bowel movements, changes in frequency and consistency of the stool [5].

An important factor in improving the diagnosis of IBS is to take into account the pathogenetic factors of the disease. In recent decades, perceptions of the pathogenesis of IBS have changed significantly. Previously, IBS was considered exclusively as a psychosomatic disease, and in almost all patients it was associated with the influence of psycho-emotional factors, but today the multifactorial development of IBS is obvious. Food allergies, stress, intes-

tinal infections, hereditary predisposition, malabsorption, and disorders of bile acid metabolism are the major triggers for the development of IBS [6]. Bile acids are amphipathic, detergent molecules synthesized by the liver that facilitate the absorption of lipids and fat soluble vitamins in the small intestine. Lithocholic and deoxycholic acids are the main bile acids present in the colon and feces. Henodeoxycholic and deoxycholic acids are known secretory bile acids. Increased excretion of feces and changes in the proportion of various bile acids in the feces characterize malabsorption of bile acids, which leads to diarrhea or IBS with diarrhea, which are associated with increased secretion of water and mucus in the colon, motility of the colon and membrane permeability. Bile malabsorption is known in 10–33% of patients with IBS with diarrhea or functional diarrhea [7].

However, the mechanisms of the link between metabolic regulation of bile acids and the pathogenesis of IBS remain unclear. Thus, studies that help identify specific pathogenetic mechanisms for the development of IBS are relevant.

THE AIM

The aim is to investigate the effect of bile acids on the ATPase activity of the colon mucosa in patients with overweight and irritable bowel syndrome.

MATERIALS AND METHODS

All procedures with patient were performed in accordance with the informed consent of the patient “International Convention for Working with Animals” under approval of the Bioethics Committee of DanyloHalytskyLviv National Medical University, protocol No2, 15/02, 2016.

Complex examination of 12 patients with IBS and excess body weight (mean age – $32,7 \pm 1,5$ years). The diagnosis of IBS was established according to Rome criteria IV [5] in the presence of recurrent abdominal pain, which was observed at least 1 day per week for the last 3 months and when there were two or more of the following symptoms: abdominal pain associated with bowel movements, pain accompanied by changing the frequency of stools or form of feces. For diagnosis of inflammatory bowel pathology, CITO TEST Calprotectin-Lactoferrin (Pharmasco) was performed. We payed attention to the absence of symptoms of anxiety: fever, impurities of blood in the stool, intestinal disorders, weight loss for a short period of time, anemia, leukocytosis, acceleration of erythrocyte sedimentation rate. All patients performed measurements of height and body weight. Body mass index was calculated by the Kettle formula. According to the obtained indicators, we established the presence of excess body weight.

Isolation of subcellular postmitochondrial fraction of the patients' colon mucous. Tissue samples were collected from patients colon during colonoscopy. Fresh samples were washed by medium A (mM): sucrose – 250, ethylene glycol tetraacetic acid (EGTA) – 1, HEPES – 10; KH_2PO_4 – 1; pH 7.2. Then these samples were homogenized with glass-glass homogenizer at 300 rev/min for 10 min at 0–2 °C.

The homogenate was centrifuged for 10 min at 3.000 g using Jouan MR 1812 centrifuge (Jouan, France) to precipitate nuclei, large cells fragments, and undestroyed cells while mitochondria remained in the supernatant 1. Next centrifugation of this supernatant 1 carried out for 10 min at 8.500 g (0–2°C). After mitochondria sedimentation, supernatant 2 was used for while ATPase activity assay. To prove a membranes presence in the post-mitochondrial fraction it was sediment for 20 min at 15.000 g.

Assay of ATPase activity. ATPase activity was determined according to the content of orthophosphate that was released after ATP hydrolysis [8,9]. At the beginning of the experiment 200 μlof post-mitochondrial subcellular fraction of patients' colon mucous was transferred to a standard incubation medium containing (mM) NaCl – 50.0; KCl – 100.0; Tris-HCl – 20.0; MgCl_2 – 3.0; CaCl_2 – 0.01; pH 7.4 at 37 °C. The reaction was started by adding 3 mM ATP (Sigma, USA). Samples were incubated for 15 min at 37 °C with moderate shaking in a water bath. Before the end of incubation 0.4 ml of medium was taken for the determination of protein content by Lowry [10]. Reaction was stopped by adding 5 ml of 10% trichloroacetic acid to samples and incubating them for 30 min followed by 10 min centrifugation at 1600 g. Supernatant obtained was used to determine the content of inorganic phosphorus by the spectrophotometric method of Fiske-Subbarow [11]. We used TLC-S (Sigma, USA) at concentration 50 $\mu\text{mol/L}$ for estimating their effect on ATPase activity.

Calculation of ATPase activity. The total ATPase activity of post-mitochondrial fraction of colon mucous was calculated by the difference of inorganic phosphorus in the medias with different composition (supplemented with TLC-S – “experiment” or not supplemented – “control”) expressed as micromoles of inorganic phosphorus equivalent to 1 mg of protein per 1 h. Specific Na^+/K^+ -ATPase activity was calculated as difference of inorganic phosphorus content in medium with or without ouabain (1 mM). For the determination of $\text{Ca}^{2+}/\text{Mg}^{2+}$ -ATPase activity, the difference between the total $\text{Ca}^{2+}/\text{Mg}^{2+}$ - and Na^+/K^+ -ATPase activity was quantified. Thapsigargin was used to calculate SERCA contribution into the total $\text{Ca}^{2+}/\text{Mg}^{2+}$ -ATPase activity. Specific basal Mg^{2+} -ATPase activity was determined in incubation medium that contained 1 mM EGTA and lacked ouabain. In all experiments, incubation medium was as a control for the enzymatic ATP hydrolysis.

Data analysis. The significance of differences between experimental groups was calculated using Wilcoxon-Mann-Whitney, when a data distributions were not normal. $P \leq 0.05$ was considered to be statistically significant.

RESULTS

It was found that Na^+/K^+ -ATPase activity of subcellular fraction of colon mucous ranged from 2.32 to 15.76 and averaged (6.06 ± 1.61) $\mu\text{mol Pi/ mg protein per hour}$. TLC-S caused ranging of Na^+/K^+ -ATPase activity from 0.74 to 13.99 and averaged (7.62 ± 1.64) $\mu\text{mol Pi/ mg protein per hour}$. Therefore, no statistically significant changes were found by

bile acid on the activity of $\text{Na}^+/\text{K}^+-\text{ATPase}$ of the subcellular fraction of the colon mucous of patients with IBS.

We observed that the $\text{Ca}^{2+}-\text{ATPase}$ activity of EPR was ranging from 0.28 to 14.14. It was equal in average (5.88 ± 1.19) $\mu\text{mol Pi/ mg protein per hour}$. TLC-S adding to the incubation medium resulted in fluctuations its activity from 0.23 to 10.89 and averaged (6.51 ± 1.20) $\mu\text{mol Pi/ mg protein per hour}$. It was found that $\text{Ca}^{2+}-\text{ATPase}$ activity of PM in control ranged from 4.84 to 15.34 and averaged (8.86 ± 1.56) $\mu\text{mol Pi/ mg protein per hour}$. When TLC-S was added to the incubation medium, the activity rates of this pump ranged from 0.61 to 10.49 and averaged (6.16 ± 1.34) $\mu\text{mol Pi/ mg protein per hour}$.

We found that basal $\text{Mg}^{2+}-\text{ATPase}$ activity in postmitochondrial subcellular fractions of colon mucous of the patients with IBS ranged from 0.42 to 9.24, which averaged (6.44 ± 2.02) $\mu\text{mol Pi/ mg protein per hour}$. Addition of TLC-S to the incubation medium resulted in fluctuations in the activity of basal $\text{Mg}^{2+}-\text{ATPase}$ activity in the range from 5.16 to 32.6 and averaged (23.19 ± 5.22) $\mu\text{mol Pi/ mg protein per hour}$.

DISCUSSION

Influence of TLC-S on $\text{Na}^+/\text{K}^+-\text{ATPase}$ activity in postmitochondrial subcellular fraction of colon mucous of the patients with IBS. As $\text{Na}^+/\text{K}^+-\text{ATPase}$ plays an important role in electrolyte, water and nutrient transport across the intestinal epithelia, it is expected that the any changes in $\text{Na}^+/\text{K}^+-\text{ATPase}$ activity may have a major impact in intestinal function, namely absorption and secretion. It was shown that activities of $\text{Na}^+/\text{K}^+-\text{ATPase}$ was increased in children with toddler diarrhea, but $\text{Na}^+/\text{K}^+-\text{ATPase}$ activity was reduced in the jejuna mucosa of patients with active celiac disease [12]. So the role of activities of $\text{Na}^+/\text{K}^+-\text{ATPase}$ in IBS pathology still unknown. It is consider that perturbed *bile acid* metabolism plays a causal role in IBS [13]. It is possible to suppose that TLC-S might effect on activity of $\text{Na}^+/\text{K}^+-\text{ATPase}$ in postmitochondrial subcellular fraction of colon mucous of the patients with IBS. But we did not found the effect of TLC-S on the activity of $\text{Na}^+/\text{K}^+-\text{ATPase}$ of the subcellular fraction of the mucous membrane of the colon in patients with IBS. Our results are agreed with Hafkenschaid, who found that “the taurine derivatives TC, TCDC and TDC did not influence or even enhanced the $\text{Na}^+/\text{K}^+-\text{ATPase}$ activity” [14].

Influence of TLC-S on total $\text{Ca}^{2+}-\text{ATPases}$ activity in postmitochondrial subcellular fraction of colon mucous of the patients with IBS. The extracellular Ca^{2+} influx is balanced by Ca^{2+} released from the cytosol by both plasma membranes and the internal $\text{Ca}^{2+}-\text{ATPases}$. The total $\text{Ca}^{2+}-\text{ATPases}$ activity of the subcellular fraction consists of EPR $\text{Ca}^{2+}-\text{ATPase}$ and plasma membrane (PM) Ca^{2+} pump. EPR $\text{Ca}^{2+}-\text{ATPase}$ play an essential role in the transport of Ca^{2+} to the EPR to replenish the calcium store, promote folding and protein maturation, lipid and steroid synthesis. It is known that TLC, as well as TLC-S, mobilizes Ca^{2+} from the intracellular pool. Thus, the main effect of TLC-S is associated with an increase in calcium cells and depletion of calcium stores. Therefore, TLC-S should affect the ac-

tivity of $\text{Ca}^{2+}-\text{ATPases}$ of the subcellular fraction of colon mucous too. But we did not observe the influence of TLC-S on $\text{Ca}^{2+}-\text{ATPase}$ activity of the subcellular fraction of the colon mucous membrane of patients with IBS.

Influence of TLC-S on basal $\text{Mg}^{2+}-\text{ATPase}$ activity in postmitochondrial subcellular fractions of colon mucous of the patients with IBS. It should to note that activity of basal $\text{Mg}^{2+}-\text{ATPase}$ activity is coupled to H^+ -translocation in PM [15,16] as well as in endosomal fraction [17]. Also in hepatocytes $\text{Mg}^{2+}-\text{ATPase}$ activity is considered as markers of canalicular membrane [18]. Mg^{2+} -activated ATPase of rat colon was studied in mucosa by J.Schreiner and coauthors & Hafkenschaidin [14, 18] and in muscle layer by Kaplia 2017 [19]. It was shown that all bile acids except cholic acid, taurocholic acid and chenodeoxycholic acid depressed the $\text{Mg}^{2+}-\text{ATPase}$ activity in rat colon mucosa [14].

We found a statistically significant increasing of the activity of basal $\text{Mg}^{2+}-\text{ATPase}$ activity in subcellular fraction of colon mucous under the action of TLC-S compared with the control by 3.6 times. The obtained results by the effects of TLC-S are in full agreement with the previously observed the effect of TLC-S on the activity of basal $\text{Mg}^{2+}-\text{ATPase}$ activity in the subcellular fraction of rat liver [20].

It has been suggested that activation of basal $\text{Mg}^{2+}-\text{ATPase}$ under the action of TLC-S may indicates to the role of the endo-lysosomal system, the so-called acid store of colon mucous of the patients in developing of pathology IBS.

CONCLUSIONS

TLC-S (50 μM) increased basal $\text{Mg}^{2+}-\text{ATPase}$ in the postmitochondrial fraction of colon mucous of the patients with overweight and IBS, but had no effect on $\text{Na}^+/\text{K}^+-\text{ATPase}$ and total $\text{Ca}^{2+}-\text{ATPases}$ activity.

REFERENCES

1. Bychkov M.A., Ferents I.M. Features of the course of irritable bowel syndrome in patients with excess body weight. *Wiad Lek.* 2018; 71(3 pt 2):688-690.
2. Mishchuk V.G., Grigoruk G.V. Types of eating behavior and serotonin levels in patients with irritable bowel syndrome and constipation on the background of obesity. *J. Clin. Exp. Med. Res.* 2018;6(1):176–182.
3. Larussa T, Rossi M, Surari E et al. Use of Complementary and Alternative Medicine by Patients with Irritable Bowel Syndrome According to the Roma IV Criteria: A Single-Center Italian Survey. *Medicina (Kaunas)*. 2019; 55(2):46. doi: 10.3390.
4. Moayyedi P, Mearin F, Azpiroz F, Andersen V. Irritable bowel syndrome diagnosis and management: A simplified algorithm for clinical practice. *United European Gastroenterology Journal.* 2017; 5(6):773-88.
5. Lacy BE, Patel NK. Rome Criteria and a Diagnostic Approach to Irritable Bowel Syndrome. *J. Clin Med.* 2017; 6(11):99.
6. Stepanov Yu.M., Budzakl.Ya., Klenina I.A. Short-chain fatty acids: the role in the development of irritable bowel syndrome. *Gastroenterologia.* 2019;53(1):49-53.
7. Vijayvargiya P., Busciglio I., Burton D. Bile Acid Deficiency in Subgroup of Patients With Irritable Bowel Syndrome With Constipation Based on Biomarkers in Serum and Fecal Samples. *Clin Gastroenterol Hepatol.* 2018; 16(4): 522–527.

8. Bychkova S, Hreniuh V. Activity of ATPases in postmitochondrial fraction of lymphoma NK/Ly cells under bafilomicine and NAADP presence mitochondria. *Studia Biologica*. 2016;9(2):31–38.
9. Hreniukh V, Bychkova S, Kulachkovsky O, Babsky A Effect of bafilomycin and NAADP on membrane-associated ATPases and respiration of isolated mitochondria of the murine Nemeth-Kellner lymphoma. *Cell biochemistry and function*. 2016; 34 (8): 579-587.
10. Lowry O.H., Rosebrough N.J., Farr A.L. et al. Protein measurement with the folin phenol reagent. *J. Biol. Chem.* 1951; 193:265–275.
11. Fiske C, Subbarow Y. The colorimetric determination of phosphorus. *J Biol Chem.* 1925;66:375–400.
12. Tripp J, Muller D, Harries J Mucosal (Na⁺-K⁺)-ATPase and Adenylate Cyclase Activities in Children with Toddler Diarrhea and the Postenteritis Syndrome. *Pediat. Res.* 1980; 14: 1382-1386.
13. Hofmann A.F. Causal Role of Bile Acids in Irritable Bowel Syndrome–Constipation. *Clinical Gastroenterology and Hepatology*. 2019; 17 (1): 213–214.
14. Hafkenschied JCM Influence of bile acids on the (Na⁺-K⁺)-activated- and Mg²⁺-activated ATPase of rat colon. *Pflügers Archiv*. 1977; 369 (3): 203–206.
15. Kosterin SO, Veklich TO, Prylutsky YI. Kinetic interpretation of pH dependence of enzymatic activity of “basal” Mg²⁺-ATPase of sarcolemma of smooth muscle. *Ukr. biochem. journal*. 2005; 77 (6): 37–45.
16. Luu-The V., Goffeau A., Thinès-Sempoux D. Rat liver plasma membrane Ca²⁺- or Mg²⁺-activated ATPase. Evidence for proton movement in reconstituted vesicles. *Biochim. Biophys. Acta*. 1987; 904 (2): 251–258.
17. Saermark T., Flint N., Evans W. H. Hepatic endosome fractions contain an ATP-driven proton pump. *Biochem. J.* 1985; 225: 51–58.
18. Schreiner J., Nell G., Loeschke K. Effect of Diphenolic Laxatives on Na⁺-K⁺-activated ATPase and Cyclic Nucleotide Content of Rat Colon Mucosa in Vivo. *Naunyn-Schmiedeberg's Arch Pharmacol.* 1980; 313 (3): 249-55.
19. Kaplia A.A. Different sensitivity of Na⁺, K⁺-ATPase and Mg²⁺-ATPase to ethanol and arachidonic acid in rat colon smooth muscle under pretreatment of cellular membranes with Ds-Na. *Ukr. Biochem. J.* 2017; 89 (2): 70-77.
20. Bychkova S. Influence of tauroolithocholate 3-sulphate on activity of Na⁺, K⁺-ATPase, Ca²⁺-ATPase and basal Mg²⁺-ATPase in rat liver subcellular fraction // *Visnyk of the Lviv University. Series Biology*. 2016; 72: 194–201.

Fragment of the research work: “Pathology of the respiratory, cardiovascular and digestive systems in patients with diabetes and obesity: features of pathogenesis, diagnosis and treatment”. №: IH.09.0001.16

ORCID and contributionship:

*Iryna M. Ferents – 0000-0001-5032-2698^{B C D}
Solomiia V. Bychkova – 0000-0002-5107-3352^E
Mykola A. Bychkov – 0000-0001-6620-1751^{A, F}*

Conflict of interest:

The Authors declare no conflict of interest.

CORRESPONDING AUTHOR

Mykola A. Bychkov

Str. Yaroslava Mudrogo, 22/5, 79016, Lviv, Ukraine
tel: +3805603753044
e-mail: koloboc2000@gmail.com

Received: 17.01.2020

Accepted: 05.03.2020

A – Work concept and design, **B** – Data collection and analysis, **C** – Responsibility for statistical analysis, **D** – Writing the article, **E** – Critical review, **F** – Final approval of the article