INTRODUCTION

The impact of low temperatures on skin is widespread in medical practice and has a long history. The cryosurgical method is well-established in the treatment of various benign, precancerous and malignant skin lesions. The benefits of cryosurgery include high success rates, minor side effects, relative ease of implementation and reasonable cost [1]. In this case, cold wounds always arise after cryosurgical treatment. It is one of the long-studied issues, which until now has been comprehensively investigated and continues to be relevant. Experimental and clinical studies in this direction indicate that the results of cold skin wounds treatment cannot be considered optimally effective today. In this regard, the search for new, effective methods and means of treating cold skin wounds remains appropriate and relevant [2]. The effectiveness of wounds treatment is known to be determined by the qualitative and quantitative morphological characteristics of the formed regenerate, the features and duration of the wound process phases [2].

Some scientists emphasize that the most prominent effects of fullerene C60 are protection from radiation-in
duced injury, neuroprotection, drug and gene delivery, anticancer therapy, adjuvant within different treatments, photosensitizing, sonosensitizing, bone reparation and biosensing [9].

There are few publications devoted to the study of C60 fullerenes effect on regenerative processes in bone tissue [10] and in burn wounds of the skin [11, 12], the results of which, at times, are contradictory and are presented in the form of hypotheses. Given the unique properties of C60 fullerenes, the study of their possible use in improving the results of wounds treatment after cryodestruction of the skin looks promising. However, there are no data on the effect of C60 fullerenes on the healing processes of cold skin wounds in the scientific literature, which undoubtedly actualizes such studies.

THE AIM
The purpose of the study is to evaluate in an experiment the morphological state of a cold skin wound using an aqueous colloidal solution of fullerene C60.

MATERIALS AND METHODS
The studies were conducted at the Institute for Problems of Cryobiology and Cryomedicine of the National Academy of Sciences of Ukraine on 6-month-old hairless male rats. The experiments were carried out according to the regulations approved by the Bioethics Committee of the Institute for Problems of Cryobiology and Cryomedicine of the National Academy of Sciences of Ukraine, developed in accordance with the «General principles of experiments on animals» approved by the Third National Congress of Bioethics (Kyiv, Ukraine, 2007) and agreed with the provisions of the European Convention on the protection of vertebrate animals used for experimental and other scientific purposes (Strasbourg, France, 1986).

During the study, three groups of 10 animals were formed: group I included rats that did not undergo manipulations; animals in groups II and III underwent to cryodestructing on the lateral surface of the thigh with an instrument that allowed to maintain the temperature of the working surface of the applicator during the operation not higher than –50 °C [13]. The diameter of the cryoeffect was 10 mm. The cryoprescription time was 120 s. After cryodestruction rats in group II were injected saline solution into the abdominal cavity during a morphometric study we determined thickness of proteolyses enzymes of neutrophils [15].

RESULTS AND DISCUSSIONS
When studying microspecimens in groups II and III on days 7, 14 and 21, a wound defect with a characteristic layer of granulation tissue. Secondary necrosis, demarcation leukocyte shaft, and a layer of granulation tissue. The statistical analysis was performed the Statistica 6.0 and Microsoft Excel 2003 software package. Nonparametric methods were used to compare the parameters (Mann-Whitney U-test, Kruskal-Wallis test). The significance of the differences between the average values of the indices in the groups was taken at a significance level of p <0.05.

In groups II and III, the entire surface of the wound defect was represented by necrotic fragments of the epidermis, dermis and hypodermis, which were components of the primary necrosis area. Currently, it is believed that occurrence of this area is associated with direct destruction of cells under the influence of low temperature [14]. There is still no clear concept of the mechanism of primary damage to biological tissues during cryodestruction and the occurrence of a zone of primary necrosis. Many cryosurgeons traditionally hold a theory of P. Mazur on the two-phase mechanism of cryodestruction, according to which tissue destruction is caused by intracellular and extracellular crystallization of water, followed by its recrystallization, due to which cell membranes are damaged. Then destruction occurs, followed by cell necrosis [14]. However, since this theory was proposed to freeze cell suspensions, it does not take into account the characteristic features of biological tissue as a complex system of cells and extracellular matrix with unique thermophysical characteristics due to microcirculation and metabolic processes.

Neutrophilic leukocytes, infiltrating the tissues around the primary necrosis area, formed an inflammatory leukocyte shaft or a demarcation area after skin cryodestruction in the wound (Fig. 1). White blood cells are known to appear in the wound just a few minutes after the damage, activate the complement system, interacting with the kallikrein-kinin system, coagulation and fibrinolysis systems, and arachidonic acid derivatives. Partial lyses of a blood clot, bacterial flora, foreign bodies, and tissue detritus occurs in the area of damage under the influence of proteolyses enzymes of neutrophils [15].
An area of secondary necrosis was revealed behind the demarcation leukocyte shaft (Fig. 1), represented by necrotic altered fragments of the dermis and hypodermis. It is believed that inflammation reactions occur after the initial low-temperature damage in the tissues. Enzymes, including proteolytic ones, are released from the cells, lipid peroxidation processes are activated and antioxidant defense weakens, active oxygen forms in the tissues, incompletely oxidized metabolic products are formed and acidosis develops. Excessive formation of aggressive products is one of the causes of the emergence and spread of the secondary necrosis area [12]. In addition, development of this area is also associated with hemodynamic disorders [14].

Thus, we can conclude that the contact of the skin with the cryotool applicator was accompanied by dynamic, interconnected changes in temperature fields on the skin surface and in the underlying tissues. Ice front occurred and spread in the tissues. It can be called the main etiological factor in the development of primary tissue necrosis. In areas where the temperature of the tissues did not fall below the cryostability point of the cells, their death, apparently, is due to a wide range of metabolic disorders caused by the indirect effect of cooling on cells functioning. The main etiological factors of such disorders include ischemic disorders caused by thrombosis of blood vessels of the microvasculature, and the inflammatory reaction that develops after warming the tissues.

Behind the zone of secondary necrosis we found a layer of granulation tissue (fig. 1) with vascular, cellular and fibrous components. Their ratio varied, indicating various degrees of maturity at different periods of the experiment.

When analyzing the obtained morphometric parameters in groups II, III with an increase in the duration of the experiment, a significant (p<0.05) decrease in the thickness of the primary and secondary necrosis area, torus demarcation of leukocytes, and an increase in the thickness of the granulation tissue layer were noted (table 1).

Hereby, the results of survey microscopy and a morphometric study indicate that the healing of cold skin wounds in groups II, III is a complex, dynamic process consisting of characteristic stages (hemostasis, inflammation, proliferation, remodeling), which do not have clearly defined time borders.

In a comparative analysis, it was noted that the healing processes in group III compared with group II at all periods of the experiment were more pronounced and active, as evidenced by the results of the survey microscopy and morphometric studies.

Comparing survey microscopy in group III with group II we observed more intensive cleansing of the wound from necrotic tissue; more active proliferative processes in it with the formation of granulation tissue, characterized by an intensive rate of maturation; in the adjacent wound defect tissues of the dermis, hypodermis with underlying muscle tissue less pronounced hemodynamic disturbances and cell infiltration, characterized by the presence of leukocytes, macrophages, lymphocytes, fibroblast cells. It is known that all reparative processes occur against the background of

Fig. 1. Group III. 14 days. Structure of wound defect: fragments of necrotic tissue (zone of primary necrosis), the demarcation leukocyte shaft, a layer of necrotic tissue (zone of secondary necrosis), a layer of granulation tissue. Stained with hematoxylin and eosin, × 100.

Fig. 2. Group III. 21 days. Complete epithelization of the regenerate zone. The epithelial layer forms superficial and deep acanthotic growths. Stained with picrofuchsin according to van Gieson, × 40.

Fig. 3. Group III. 21 days. Increased mitotic activity and mild dysplastic changes in epidermis. Stained with hematoxylin and eosin, × 400.
an inflammatory reaction, which, from the morphological point of view, is divided into leukocyte, macrophage and fibroblastic stages [16]. In a morphometric study in group III compared with group II at all periods of the experiment a significantly (p<0.05) lower value of the thickness of the primary and secondary necrosis area was noted, significantly (p<0.05) a larger value of the thickness of the zone of primary necrosis and of the zone of secondary necrosis (Table 1). The more pronounced regenerative processes in group III compared with group II, were also indicated by the reduction in the width of the mitotic activity area. Proliferation intensity of the epithelium of the wound edges, but after 24 hours the mitotic index and the index of labeled nuclei increase several times. Interestingly, the size of the wound does not affect the width of the mitotic activity area. Proliferation intensity of the epithelium in the area of skin defects is variable, it is possible to develop severe dysplasia, the combination of which with enhanced proliferation is a characteristic feature of precancerous skin diseases. In order to refute or confirm these fears, it is necessary to study proliferative processes in epithelium with the aim of their qualitative assessment.

Initially, a decrease in the synthesis of deoxyribonucleoproteins and the number of mitoses is noted in the damaged epithelium of the wound edges, but after 24 hours the mitotic index and the index of labeled nuclei increase several times. Interestingly, the size of the wound does not affect the width of the mitotic activity area. Proliferation intensity of the epithelium in the area of skin defects is variable, it changes, increasing and decreasing at certain intervals. It is known that the proliferative activity of the epithelium can be inflammatory or regenerative in nature [17]. Hypertrophic and hyperplastic processes were also detect-

### Table 1. Thickness of layers (×10-6m) of wound defect in groups II and III.

<table>
<thead>
<tr>
<th>Layer name</th>
<th>Term of animals withdrawn from experiment, days</th>
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<tbody>
<tr>
<td></td>
<td>7</td>
</tr>
<tr>
<td>zone of primary necrosis</td>
<td>1,975.9±93.7</td>
</tr>
<tr>
<td></td>
<td>p&lt;0.05</td>
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<tr>
<td>leukocyte demarcation shaft</td>
<td>329.5±33.2</td>
</tr>
<tr>
<td></td>
<td>p&lt;0.05</td>
</tr>
<tr>
<td>zone of secondary necrosis</td>
<td>482.7±19.0</td>
</tr>
<tr>
<td></td>
<td>p&lt;0.05</td>
</tr>
<tr>
<td>granulation tissue layer</td>
<td>148.3±20.4</td>
</tr>
<tr>
<td></td>
<td>p&lt;0.05</td>
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</tbody>
</table>

Note: p₁ – significance of differences compared with previous term of the animals withdrawn from experiment, p₂ – significance of differences compared with group II.

### Table 2. Thickness of the epitelial layer (×10-6m) in groups I-III.

<table>
<thead>
<tr>
<th>Group number</th>
<th>The term of the animals withdrawn from experiment, days</th>
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<tbody>
<tr>
<td></td>
<td>7</td>
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<tr>
<td>I</td>
<td>(21.3±0.7) ×10⁻⁶m</td>
</tr>
<tr>
<td></td>
<td>p&lt;0.05</td>
</tr>
<tr>
<td>II</td>
<td>(26.8±1.2) ×10⁻⁶m</td>
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<tr>
<td></td>
<td>p&lt;0.05</td>
</tr>
<tr>
<td>III</td>
<td>(69.9±2.1) ×10⁻⁶m</td>
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<td></td>
<td>p&lt;0.05</td>
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Note: p₁ – significance of differences compared with group I, p₂ – the significance of differences compared to previous period of animal withdrawal from the experiment, p₃ – the significance of the differences compared with group II.
ed in the above areas in the epidermis, which has led to a significant (p<0.05) increase in the thickness of this layer in groups II and III as compared with group I (table 2). With an increase in the experiment duration the thickness of the epithelial layer in groups II and III increased significantly (p<0.05), indicating an increase in regenerative processes. This index was characterized by a significantly (p<0.05) higher value in group III compared with group II.

Re-epithelialization of the wound [18] begins with the migration of epithelial cells from the edges of the wound to the area of the tissue defect within a few hours after tissue damage, continuing throughout all phases of the wound healing. Weakening of intercellular contacts and cell contacts with the basement membrane, formation of peripheral cytoplasmic actin filaments make it possible for epithelial cells to move in the direction of the damaged tissues. As the process of re-epithelialization advances, the basal membrane is re-formed [15].

Keratinocytes adjacent to the lesion area also affect the course of wound healing. Various isoforms of transforming growth factor β synthesized by them, as well as platelet growth factor, affect the proliferation of fibroblasts, their migration to the area of damage, and production of extracellular matrix components [15].

We have discovered the enhancement of regenerative potentials in the wound after cryodestruction of the skin in hairless rats, which were injected into the abdominal cavity with an aqueous colloidal solution of C60 fullerenes. From our point of view, it is firstly, due, to the antioxidant activity of fullerenes [12], which prevents damage to cells and tissues by secondary alteration products; secondly, the ability of fullerenes to induce differentiation of fibroblasts [19], which, apparently, leads to the activation of collagenogenesis, rapid filling of a wound defect with granulation tissue with its intensive maturation and transformation into connective tissue.

CONCLUSIONS

A comprehensive morphological study of the experimental material indicates that the abdominal cavity injection of an aqueous colloidal solution of C60 fullerenes activates reparative processes in the skin cold wound, going through all the classical stages.

C60 fullerenes stimulate proliferative activity in the epidermis, located in the wound marginal regions or covering the regeneration surface, which leads to an increase in epithelial layer thickness by 2.6, 2.6, 2.5 times on 7, 14 and 21 days; promotes faster cleaning of the wound from necrotic tissue, reducing thickness of primary necrosis area on days 7, 14 and 21 by 1.3 times by increasing the demarcation area on days 7, 14 and 21 by 1.4, 1.4 and 2, 2 times; reduces the thickness of the secondary necrosis area by 7, 14 and 21 days by 1.4, 1.5 and 1.3 times; accelerates the filling of the wound defect with granulation tissue, the layer thickness of which increases by 2.3, 2.2 and 1.4 times on the 7th, 14th and 21st days, respectively; reduces the severity of hemodynamic disorders and cell infiltration in the wound defect adjacent tissues of the dermis, hypoder-mis with the underlying muscle tissue.

The prospect of further research is an immunohistochemical investigation to discover the apoptotic and prolif-erative processes in the wound after skin cryodestruction in hairless rats.

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Authors declare no conflict of interest.

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